Protective effects of spironolactone against hepatic ischemia/reperfusion injury in rats

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ABSTRACT

Objective: In the present study, it was aimed to study the antioxidant effects of spironolactone (SPL) to determine its possible protective effects in hepatic ischemia reperfusion injury.

Material and Methods: Hepatic artery, portal vein, and bile duct of Wistar albino rats were clamped for 45 minutes under anesthesia to form an ischemia period. Then reperfusion was allowed and the rats were decapitated 60 minutes later. SPL (20 mg/kg, p.o.) or SF was orally administered for 30 minutes before ischemia. Rats in the control arm underwent sham surgery and were administered isotonic saline. Liver function was studied by measuring aspartate aminotransferase (AST), alanine aminotransferase (ALT), tumor necrosis factor-alpha (TNF-α), and interleukin 1beta (IL-1β) levels. Malondialdehyde (MDA), glutathione (GSH), luminol, and lucigenin levels, myeloperoxidase (MPO) and Na+K+-ATPase enzyme activities were analyzed to study tissue injury under light microscope.

Results: While IR increased AST, ALT, TNF-α, and IL-1β levels and MDA, luminol, and lucigenin levels and MPO activities, it caused a decrease in GSH levels and Na+K+-ATPase activity. Spironolactone administration significantly improved these values.

Conclusion: Protective effects of SPL against ischemia/reperfusion injury via various mechanisms suggest that this agent may become a novel treatment agent in clinical practice.

Keywords: Hepatic ischemia reperfusion, spironolactone, malondialdehyde, glutathione, cytokines

INTRODUCTION

Ischemia/reperfusion injury takes place in various clinical settings such as hepatic trauma, hemorrhagic shock, resection of large intrahepatic tumors, and liver transplantation (1,2). The pathophysiology of hepatic ischemia reperfusion injury involves many mechanisms. It consists of several major steps including Kupffer cell activation, formation of reactive oxygen derivatives, cytokine release, activation of polymorphonuclear leukocytes, altered mitochondrial permeability, and activation of endothelial cells and complement systems, among several other cellular and molecular mechanisms (3,4). These events lead to the activation of local and systemic inflammatory responses, causing injury to local organs.

Aldosterone is secreted by zona glomerulosa in the adrenal cortex. Spironolactone is a specific aldosterone antagonist. It acts by binding to aldosterone receptors, sparing potassium ions while removing sodium and water in distal tubuli. Its impact on renal ischemia/reperfusion injury was demonstrated in a study conducted by Meija-Vilet et al (5). To date, however, no researcher has studied the effects of spironolactone on hepatic ischemia/reperfusion. We therefore consider that our work will have an important place in the literature by being the first study that investigates this subject.

Under the light of the above mentioned information, this study aimed to investigate the effects of spironolactone on liver injury resulting from ischemia/reperfusion...
applied at the level of middle and left hepatic artery, portal vein, and bile ducts.

**MATERIAL and METHODS**

This experiment was performed after obtaining the approval of the Local Ethics Committee at Marmara University Laboratory Animals Research Center with the protocol code numbered 80.2012.mar, dated 08.11.2012. The rats were obtained from and the experiment was conducted at the same laboratory.

The study used 32 ten-week-old female Norwegian rats of the Wistar albino strain weighing between 200 and 250 grams. The rats were randomly grouped into 4 groups containing 8 rats each and they were kept in wire cages containing 4 rats each under standard laboratory conditions (20-24°C, 50-60% humidity; 12-hour cycles of light and darkness); the rats were fed ad libitum with standard rat feed and tap water during the whole study period. No enteral or parenteral antibiotics were administered throughout the study.

Each rat was kept in a jar containing ether for 40-60 seconds for anesthesia induction. Following anesthesia induction, ketamin at 100 mg/kg and chlorpromazine at 1 mg/kg were administered intraperitoneally (ip) for anesthesia maintenance.

The rats were then laid supine under anesthesia. The anterior abdominal wall was shaved with a scalpel and the shaved area was sterilized with povidone iodine solution. Then, the abdominal cavity was entered via midline incision. Hepatic artery, portal vein, and bile duct were explored and clamped for 45 minutes to create ischemia. At the end of this period, reperfusion was established, and the rats were decapitated 60 minutes later. Spironolactone (20 mg/kg, p.o) or isotonic saline were applied via oral route for 30 minutes before ischemia. The rats in the control group were applied sham operation and isotonic saline infusion.

1. **Sham group:** Following the completion of the standard operation, a 2-cc blood sample and a hepatic tissue sample were taken without clamping the hepatic artery, portal vein, and bile ducts.

2. **Spironolactone (SPL) group:** The rats were orally administered SPL at a dose of 20 mg/kg for 30 minutes before the time of the sham operation. Then a 2-cc blood sample and a hepatic tissue sample were taken.

3. **Ischemia reperfusion (I/R) group:** The rats were orally administered isotonic for 30 minutes before the time of the sham operation. Following the completion of the standard operation, hepatic artery, portal vein, and bile ducts were clamped to create ischemia. The rats were subjected to ischemia for only 45 minutes, followed by 60 minutes of reperfusion. At the end of the reperfusion period, a 2-cc blood sample was obtained from the heart and a tissue sample was taken from liver.

4. **The Spironolactone administered ischemia/reperfusion (I/R-SPL) group:** The rats were orally administered spironolactone at a dose of 20 mg/kg for 30 minutes before the time of the hepatic ischemia procedure. The rats were subjected to ischemia for only 45 minutes, followed by 60 minutes of reperfusion. At the end of the reperfusion period, a 2-cc blood sample was obtained from the heart and a tissue sample was taken from the liver.

**Serum Studies**

Determination of serum AST (Biolabo Europe S.A. Catalog No: REF 92025, 02160 Maizy, France) and ALT (Biolabo Europe S.A. Catalog No: REF 80027, 02160 Maizy, France) is a kinetic spectrophotometric method which was performed in the Opera Technican Bayer Autoanalyser (Germany) device. TNF-α (BioSource Europe S.A. Catalog No.KRC 3014, Nivelles, Belgium) and IL-1β (ELISA, BioSource Catalog No.KRC0011, Nivelles, Belgium) levels were determined with the ELISA method using specific rat kits.

**Tissue Studies**

Determination of free oxygen radicals in tissue was performed by the Chemiluminescence Method (6,7). Tissue samples were put into 2 ml PBS-Hepes buffer, and 4 ml luminol (0.2 mM) and 4 ml lusigenin (0.2 mM) were added to 2 separate tubes containing samples of the same tissue. Photonic activity occurring as a result of the reaction of reactive oxygen species and luminol and lusigenin were recorded by a luminometer for 10 minutes and the “area under curve” (AUC) formed by the count was calculated.

To determine MDA and GSH levels, samples of the tissue were homogenized with ice-cold 150 mM KCl. As described by Beuge et al. in 1978 (8) for products of lipid peroxidation, MDA levels were assayed by monitoring thiobarbituric acid reactive substance formation. Lipid peroxidation was expressed in terms of MDA equivalents using an extinction coefficient of 1.56 x 10⁵ M⁻¹ cm⁻¹ and results were expressed as nmol MDA/g tissue. GSH measurements were performed with the modification of the Ellman procedure (9). After centrifugation at 1077 x g for 10 min, 0.5 mL of supernatant was added to 2 mL of 0.3 mol/L Na₂HPO₄·2H₂O solution. A 0.2 mL solution of dithiobisnitrobenzoate (0.4 mg/ml 1% sodium citrate) was added, and the absorbance at 412 nm was measured immediately after mixing. GSH levels were calculated using an extinction coefficient of 1.36 x 10⁴ M⁻¹ cm⁻¹. Results were expressed in μmol GSH/g tissue.
Tissue MPO was measured with the Hillegeas method (10). Tissue samples taken immediately after decapitation were washed with isotonic saline to remove any blood clots and debris, dried with a filter paper, and weighed. The liver sample was homogenized with 50 mM K$_2$HPO$_4$ (pH: 6) to prepare a 10% homogenate, and it was centrifuged at 41400 g for 10 minutes at 4°C. The supernatant was discarded and the samples that were re-homogenized by adding 0.5% HETAB (Hexadesyltrimethyl-ammonium bromide) to the precipitate were frozen, thawed, and sonicated for three times. Then the samples were centrifuged at 41400 g for 10 minutes at 4°C. The supernatant was discarded and the precipitate was added 50 mM K$_2$HPO$_4$ (pH: 6), 20 mM H$_2$O$_2$ and o-Dianizidine-2 HCl. It was incubated in a 37°C water bath for 3 minutes, and added 2% sodium azide to stop color reaction. The absorbance level of the samples were then centrifuged at 41400 g for 10 minutes at 4°C, the supernatant was discarded, and the absorbance level of the final color was read at 460 nm in spectrophotometry.

Determination of tissue Na$^+$/K$^+$-ATPase activity was done with the Reading and Isbir method (11). The liver tissue sample was homogenized in a 10% sucrose solution and centrifuged at 3000 rpm for 10 minutes. Zero point one milliliter of the supernatant was discarded and the homogenates were incubated in a suitable medium containing 3 mM ATP. Mg$^{2+}$-ATPase activity was determined in the presence of 1 mM ouabain while total ATPase activity was determined in the presence of 100 mM NaCl, 5 mM KCl, 6 mM MgCl$_2$, 0.1 mM EDTA, and 30 mM Tris HCl (pH 7.4). The difference between total ATPase activity and Mg$^{2+}$-ATPase activity was considered Na$^+$/K$^+$-ATPase activity, and the specific activity of the enzyme was expressed as nmol Pi mg$^{-1}$ protein$h^{-1}$. The protein concentration of the supernatant was measured with Lowry method (12).

**The Method of Histopathological Examination**

**Light microscope:** The tissue samples were put into 10% formalin, washed with tap water for at least 3 hours or overnight, dehydrated with increasing alcohol concentrations (70% alcohol for 2 minutes, 90% alcohol for 2 minutes, 70% alcohol for 30 minutes, 100% alcohol twice for 30 minutes, 100% toluene twice) and put in distilled water. Following treatment with hematoxylin for 15 minutes, the samples were put in tap water for embyrulping process. After adding eosin and distilled water for 5 minutes, dehydration procedure was repeated with increasing concentrations of alcohol (70% alcohol for 2 minutes, 90% alcohol for 2 minutes, 96% alcohol for 2 minutes, 100% alcohol for 10 minutes). Then the samples were washed with Toluene twice (first bath for 5 minutes, second bath for 10 minutes), covered with Entellan, and examined under light microscope.

**Statistical Analysis**

Serum ALT, AST, TNF-α, and IL-1β levels and hepatic GSH, MDA, luminol, lucigenin levels, Bcl-2/Bax ratio, and MPO, Na$^+$, K$^+$-ATPase, caspase 3 and caspase 9 activities were compared with one-way analysis of variance (ANOVA) test, with paired comparisons being performed with Tukey’s test.

A p value of less than 0.05 was considered statistically significant.

**RESULTS**

In our study ALT, AST, TNF-α, IL-1β levels in serum, and GSH, MDA, luminol, lucigenin levels, MPO, Na$^+$, K$^+$-ATPase activities in liver tissue were compared between the groups with one-way analysis of variance (ANOVA) test, with paired comparisons being performed with Tukey’s test.

In the group applied with hepatic IR serum, AST, ALT, TNF-α, and IL-1β levels were significantly higher than the sham group. On the other hand, these parameters approximated to the control levels with SPL administration (Table 1).

Hepatic GSH level was lower in the IR group than the sham group. This decrease was largely prevented after the SPL administration in the IR group (Figure 1A). MDA levels were significantly higher in the IR group than the sham group. SPL administration significantly blunted this increase in the MDA levels (Figure 1B).

**Table 1.** Serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), tumor necrosis factor-alpha (TNF-α) and interleukin 1-beta (IL-1β) levels of the study groups in hepatic ischemia/reperfusion (I/R) model in rats.

<table>
<thead>
<tr>
<th></th>
<th>Sham</th>
<th>SPL</th>
<th>I/R</th>
<th>I/R-SPL</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/L)</td>
<td>56.5 ± 7.5</td>
<td>61.1 ± 5.5</td>
<td>139.2 ± 15.1***</td>
<td>62.3 ± 8.8***</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>53.1 ± 4.9</td>
<td>59.7 ± 5.6</td>
<td>142.5 ± 14.8**</td>
<td>90.8 ± 12.6*</td>
</tr>
<tr>
<td>TNF-α (pg/ml)</td>
<td>51.2 ± 2.96</td>
<td>44.5 ± 3.1</td>
<td>78.70 ± 4.2***</td>
<td>51.90 ± 2.2**</td>
</tr>
<tr>
<td>IL-1β (pg/ml)</td>
<td>359 ± 22</td>
<td>319 ± 16</td>
<td>466 ± 28**</td>
<td>371 ± 11*</td>
</tr>
</tbody>
</table>

Mean ± standard error.

**p< 0.01, ***p< 0.001 vs. sham group.
+ p< 0.05, +++ p< 0.001 vs. I/R group.
SPL: Spironolactone.
Hepatic luminol and lucigenin levels were significantly higher in the IR group than the sham group. This increase was observed to be significantly inhibited by SPL in the IR group (Figure 2A, B).

While hepatic ischemia led to a significantly increased neutrophil infiltration and thus MPO levels in the hepatic tissue compared to the sham group, MPO activity was significantly reduced in the SPL administered group and approached the MPO levels of the sham group (Figure 3A). Na⁺-K⁺-ATPase activity was lower in the IR group than the sham group. This reduction was largely prevented in the SPL administered group (Figure 3B).

The histopathological examination of the sham group showed that sinusoids and hepatocytes were intact and liver parenchyma preserved its normal morphology (Figure 4A). SPL administered group revealed intact sinusoids and hepatocytes with preserved morphology of the liver parenchyma (Figure 4B), IR group demonstrated extensive congestion and hepatocyte...
Ischemia, defined as hepatocyte degeneration despite mild congestion (Figure 4D). Registered IR group demonstrated low-intensity sinusoidal and degeneration caused by ischemia (Figure 4C) and SPL administered IR group showed reduced congestion in sinusoids (arrows) and central vein (*)

**DISCUSSION**

Ischemia, defined as oxygen deprivation of any tissue or organ as a result of insufficient perfusion secondary to the cessation of blood flow, and subsequent reperfusion together trigger a series of pathological reactions resulting in cell death and organ dysfunction. During the process of ischemia, cell death ensues as a result of the depletion of cellular energy stores and accumulation of toxic metabolites. Restoration of blood flow is essential for both cellular regeneration and clearance of toxic metabolites. Nevertheless, a number of studies have indicated that tissue reperfusion paradoxically gives rise to a substantially more severe injury in ischemic tissue compared to that inflicted by ischemia per se (13).

Although dysfunction following reperfusion injury shows variability depending on structural alterations of an organ, oxygen radicals have been determined to be responsible for this injury in every organ (14).

In hepatic tissue, it was first observed in an experimental liver transplantation carried out by Toledo-Pereyra et al. in 1975 (15). It resulted in graft necrosis in the transplanted liver, which culminated into congestion, progressive thrombosis, and organ failure (16). Since blood loss during major liver surgery is closely related to mortality and morbidity, it should be kept at minimum through vascular isolation of the liver. Maneuvers performed to limit afferent blood flow of the liver primarily result in hepatic ischemia. Once the obstacle to hepatic blood flow is removed, the reperfusion process begins. IR injury is the leading cause of post-transplant dysfunction (17).

In an experimental study in rats, we biochemically and immunohistochemically evaluated pathological alterations in the liver tissue secondary to hepatic IR which was formed by clamping the middle and left hepatic arteries, portal vein, and bile duct. We aimed to assess the antioxidant and antiapoptotic actions of SPL, a mineralocorticoid receptor antagonist, in an attempt to investigate its protective effects against IR injury.

Although there exist various different methods to assess liver function after ischemia reperfusion injury, determination of the activities of AST, ALT is currently the most accepted method. The activities of these enzymes are known to increase in hepatic injury (18). Various studies have shown the antioxidant protective effects of mineralocorticoid receptor antagonists (19,20).

In our study, serum AST and ALT levels were significantly higher in the hepatic IR group compared to the control group. In contrast, the levels approached the control levels upon the administration of SPL. These results suggest that tissue injury was less intense in the SPL administered group.

TNF-α is a cytokine released from monocytes, macrophages, and T cells that are abundant in the peritoneum and splanchic tissues. It is also found in Kupffer cells, the largest macrophage aggregate in the human body. TNF-α has some significant roles in muscle catabolism during stress and cachexia (1). In an experimental renal IR model, Kabasakal et al. have shown that IR injury can be reduced by inhibiting the formation of TNF-α (21). The result that we found is parallel to the literature. We found significantly higher TNF-α levels in the hepatic IR group when compared with the control group. In contrast, the SPL administered IR group showed a significantly blunted increase in TNF-α level, which approached that of the control group.

IL-1 is released from KCs in response to inflammatory stimuli. It is also synthesized by neutrophils, epithelial cells, and endothelial cells. Kato et al. have reported that IL-1β level is one of the major indicators of hepatic IR injury (22). In our study, the hepatic IR group had significantly higher serum IL-1β levels than the control group. On the other hand, SPL administered group enjoyed a significantly reduced increase, and IL-1β levels approached the control levels. Mineralocorticoid receptor antagonist Spironolactone has been shown to inhibit proinflammatory cytokines in previously conducted inflammation studies (23,24). The results that we found in our study are parallel to the literature. Because of the hepatic ischemia/reperfusion, increased values were near to normal values with spironolactone application.

Lipids are among the main targets of free radicals that are formed during ischemia reperfusion. Lipid peroxidation is considered by some researchers a key factor in IR injury (25).
Reactive oxygen species take one hydrogen atom from polyunsaturates fatty acids and initiates lipid peroxidation to form hydroperoxides. These reactions cause cell membrane to lose its liquidity and integrity, which ultimately leads to the release of cellular fractions to the surrounding medium, and cell death. These subcellular structures trigger inflammatory events and further aggravate injury (26). Different methods are used to detect tissue lipid peroxidation. One of the most widely used among them is the determination of the levels of MDA, a conjugated diene. Our study showed that MDA levels were significantly higher in the liver IR group compared to the controls. SPL administration study showed that MDA levels were significantly higher in the liver IR group as compared to the controls. SPL administration substantially reduced the increase in MDA level.

All aerobic living beings are subjected to oxidative stress during cellular metabolism. Free radicals such as \( \text{H}_2\text{O}_2 \), \( \text{O}_2 \), which are formed in an organism, are transformed into more toxic metabolites and attack various targets including DNA, lipids, and proteins, disrupting metabolic processes. However, organisms possess many defensive mechanisms to neutralize these reactive agents. Referred to as the “antioxidant defense system”, this system has glutathione as the most important component. GSH reacts with free radicals and peroxides to protect cells against oxidative injury. GSH concentration has been shown to be reduced rapidly during hepatic IR injury as a result of being used to neutralize reactive oxygen molecules. Many studies have shown that exogenously administered GSH increases intracellular GSH levels and prevents oxidative injury. Therefore, GSH has a major protective effect against oxidative injuries (27). Our study indicated lower hepatic GSH levels in the IR group than the sham group. This reduction in GSH levels was prevented by SPL in the IR group.

Studies have shown that agents used as antioxidants prevent the increase in MDA levels by increasing GSH levels (28,29). In this study, Spironolactone, which we used as a protective agent, increased the levels of decreased glutathione in the liver and significantly decreased the levels of MDA.

Neutrophils activated by ischemia reperfusion release myeloperoxidase into the medium and induce the formation of hypochloric acid (HOCI) from \( \text{H}_2\text{O}_2 \) and chloride ions. HOCI exerts cytotoxic effects by oxidizing sulphides, inactivating cytochrome and heme proteins, and degrading amino acids and proteins (30). Tissue MPO activity is considered an indicator of neutrophil accumulation. Our study demonstrated that while the induction of hepatic ischemia caused neutrophil infiltration and an associated significant increase in MPO activity compared to the control group, MPO activity was significantly lowered in the SPL administered group, approaching to that in the sham group. Considering the positive effects of Spironolactone on blood parameters used in our study, the fact that these agents significantly suppressed myeloperoxidase enzyme in the tissues supports that this enzyme plays an important role in HI. Thus, the presence of inhibitor properties on the MPO activity of the agents used in HI therapy will ensure that the tissue is protected against oxidation and will increase the success of treatment. Spironolactone has been reported to reduce increased myeloperoxidase activity during inflammatory events by the effect of free radical scavenging (31). When all these studies are taken into consideration, it is understood that our findings are in conformity with the literature.

Na⁺-K⁺-ATPase is an important membrane enzyme playing a key role in hepatocyte structure and physiology by maintaining sodium-potassium gradient on cellular membrane. It is an important indicator of maintenance of tissue viability and liver function. Studies have shown a reduction in Na⁺-K⁺-ATPase level after hepatic ischemia reperfusion (32,33). Our study revealed that Na⁺-K⁺-ATPase activity in the liver tissue was lower in the IR group when compared to the control group. This reduction was largely prevented by treatment with SPL.

Luminol hydroxyl radical is an indicator of the formation of reactive oxygen metabolites such as hydrogen peroxide and hypochloric acid. Lucigenin marks the formation of superoxide anion radical (33). They indirectly indicate the level of ROM-mediated tissue injury. In our study, luminol levels were higher in the IR group than the control group, and the increase was prevented by SPL administration. Lucigenin levels were significantly higher in the hepatic IR group compared to the controls. SPL administration, however, reduced this elevation to a significant extent. Spironolactone suggests that this effect is probably accomplished by inhibiting neutrophil infiltration.

Hepatic IR injury is known to induce hepatocellular necrosis and sinusoidal congestion. In our study, the histopathological examination of the control and SPL groups revealed that sinusoids and hepatocytes were intact, and liver parenchyma preserved its normal morphology; the histopathological examination of the IR group showed that hepatic ischemia caused extensive congestion and hepatocyte degeneration; and the histopathological examination of the IR group treated with SPL indicated a down grade sinusoidal and hepatocellular degeneration despite mild congestion.

In conclusion, it was observed that GSH level and Na⁺-K⁺-ATPase activity intensity decreased, luminol, lucigenin, MDA level and MPO activity increased, and accordingly, hepatic dysfunction occurred in the liver after IR of the hepatic tissue. SPL provided protection against IR by largely reducing IR-induced injury. These results suggest that SPL treatment can lower the morbidity and mortality of hepatic IR to a considerable extent by preventing IR-induced free radical-mediated organ injury and dysfunction.

**Ethics Committee Approval:** This experiment was performed after obtaining the approval of the Local Ethics Committee at Marmara University Laboratory Animals Research Center with the protocol code numbered 80.2012.mar. dated 08.11.2012.
**ÖZET**

**Giriş ve Amaç:** Bu çalışmada, hepatik iskemi/reperfüzyon hasarında olası koruyucu etkilerini saptamak için spironolaktonun (SPL) antioksidan etkilerinin araştırılması amaçlanmıştır.

**Gereç ve Yöntem:** Wistar albino sıçanların hepatik arter, portal ven ve safra kanalı, bir iskemi dönemi oluşturmak için anestezi altında 45 dakika boyunca klempledilmiştir. 60 dakikalık reperfüzyon sonunda sıçanlar dekapite edilmiştir. SPL (20 mg/kg, p.o.) veya SF, iskemiden 30 dakika boyunca oral yolla uygulandı. Kontrol grubuna ise sham cerrahisi ve izotonik salin verildi. Serumda aspartat aminotransferaz (AST), alanin aminotransferaz (ALT), tümör nekrozis faktör-alfa (TNF-α) ve interlökin-1beta (IL-1β) düzeyleri ölçüldü. Malondialdehid, glutatyon, luminol ve lucigenin seviyeleri, miyeloperoksidaz ve Na⁺-K⁺-ATPase enzim aktiviteleri ve ışık mikroskobu altında doku hasarını incelemek üzere analiz edilmiştir.

**Bulgular:** IR, AST, ALT, TNF-α ve IL-1ß seviyelerini ve MDA, luminol ve lucigenin seviyelerini ve MPO aktivitelerini artırırken, GSH düzeylerinde, Na⁺-K⁺-ATPase aktivitesinde azalmaya neden olmuştur. Spironolakton uygulaması bu değerleri önemli ölçüde iyileştirmiştir.

**Sonuç:** SPL'nin iskemi/reperfüzyon hasarına karşı çeşitli mekanizmalarla koruyucu etkileri, bu ajanın klinik uygulamada yeni bir tedavi ajanı olabileceğini düşündüremektedir.

**Anahtar Kelimeler:** Hepatik iskemi reperfüzyon, spironolakton, malondialdehit, glutatyon, sitokinler

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