Evaluation of the effects of adipose-derived mesenchymal stem cells on intraperitoneal adhesions

Erdal Uysal¹, Mehmet Dokur², Türkay Kırdak³, Akif Kurt⁴, Mehmet Karadağ⁵

ABSTRACT

Objectives: The goal was to examine the efficiency of local application of adipose-derived mesenchymal stem cells, which have an anti-inflammatory effect, in preventing the intra-abdominal adhesions in rats.

Material and Methods: Twenty-one Wistar albino rats were randomly divided into 3 groups, 7 rats in each: Group 1 was defined as the control group, Group 2 as the sham group, and Group 3 as the adipose-derived mesenchymal stem cell group. A 6 cm mid-abdomen incision in the all the rats was performed. The cecum serosa and sub-serosa were injured by rubbing with a gauze. No agent was applied intraperitoneally for the rats in Group 1; 1.5 ml saline and 2x10⁶/kg allojenic adipose-derived mesenchymal stem cells in the 1.5 ml saline were injected into peritoneum of rats in Groups 2 and 3, respectively. Laparotomy was performed on the 14th day. Adhesion scores, histopathological examination, E-cadherin expression, and the tissue hydroxyproline level were evaluated.

Results: The general adhesion score and collagen deposition in Group 3 were found to be significantly higher than in Groups 1 and 2 (p=0.003 and p=0.009, respectively). In the inflammatory cell comparison, a significant decrease was found in Group 3 in proportion to Groups 1 and 2 (p=0.001, p=0.005, respectively). The E-cadherin levels were found to be higher in Group 3 (p=0.003).

Conclusion: Severe adhesion was observed in the adipose-derived mesenchymal stem cells group. Collagen intensity and E-Cadherin expression also increased in the adipose-derived mesenchymal stem cells group. The anti-inflammatory effect was also seen in the adipose-derived mesenchymal stem cells group.

Keywords: E-cadherin, rat, hydroxyproline, peritoneal adhesion, stem cells

INTRODUCTION

Intraperitoneal adhesions that may develop after a great number of abdominal operations is an important problem that surgeons have to deal with after the abdominal surgical procedures. Although the incidence varies between 67% and 93%, the incidence of cases requiring surgical intervention due to intraperitoneal adhesion is 15%–18% (1, 2). The development of minimal invasive methods such as laparoscopy and their common usages decreased these rates (3). Intraperitoneal adhesions lead to hospitalization and surgical intervention due to mechanical ileus and pelvic pain. Furthermore, the intraperitoneal adhesions may cause secondary infertility in women during the fertility age (4). Since the exploration is very difficult in operations performed due to the adhesion, the iatrogenic injuries may be seen more frequently. Consequently, these complications lead to mortality and morbidity. Moreover, one of the most important results of adhesion-caused ileus is the increase in workforce loss and patient costs. For instance, in a study that has been carried out in the United States, it has been reported that the cost of surgeries performed due to the intraperitoneal adhesions was US$1.3 billion in a year (5).

To decrease and prevent the adhesions, the adhesion-barrier agents such as carboxymethylcellulose, steroids, and non-steroidal anti-inflammatory drugs, immunosuppressive drugs, clinoleic against, high molecular weight hyaluronic acid and low molecular weight hydroxypropyl methylcellulose, fibrin destruction agents, recombinant tissue plasminogen activator, bevacizumab, and Vitamin-E have been utilized (6-10). Different results have been reported for these agents. Except for the adhesion-barrier, others could not be put into the practice. Therefore, preventing the intraperitoneal adhesions and dealing with them have constantly remained on the agenda of surgeons interested in abdominal surgery.

It has been reported that post-operative inflammatory responses play a significant role in the formation of adhesions (11). The inflammatory response, especially the suppression of TGF-β1 and IL-6, may play an effective role in decreasing the intraperitoneal adhesion (12). Therefore, the anti-inflammatory feature of the selected chemical agent to prevent the peritoneal adhesion becomes very important. Adipose derived mesenchymal stem cells (ADSCs) show an anti-inflammatory effect by decreasing the inflammation in tissues (13, 14). Under the light of these observations, it can be hypothesized that ADSCs may decrease or prevent the adhesion formation by exhibiting the anti-inflammatory effect in the development of postoperative adhesions. In this study, the goal was to evaluate the effect of ADSCs on intraperitoneal adhesions that have an anti-inflammatory effect and on the intra-abdominal adhesions in rats.
MATERIAL AND METHODS
The present study was approved by the local ethic committee. The protocol was approved by the Animal Ethics Review Committee (Permit number 2016/01-02). All experiments were conducted in compliance with the relevant laws and institutional guidelines.

Rat groups
Twenty-one female Wistar albino rats weighing between 260 and 280gr were randomly divided into 3 groups, 7 rats in each: Group 1 was defined as the control group, Group 2 as Sham, and Group 3 as the ADSCs group. All rats were females to provide standardization in the study.

The cecum serosa of the rats was injured with a sterile gauze during laparotomy. For the rats in Group 1, no agent was implemented intraperitoneally. For the rats in Group 2, 1.5 ml saline was locally injected into the peritoneum. For the rats in Group 3, 2x10⁶/kg allojenic ADSCs in the 1.5 mL saline were injected locally into the peritoneum.

Anesthesia and Surgical Technique
Rats were kept between 18–24°C (12 hours of day-night cycle) in cages (4–6 rats in each) with free water and food intake. Standard rat food and water were given to the rats. Before the operation, rats were fasting for 6 hours. As the anesthetics, ketamine (Ketalar, Pfizer, Turkey) 75 mg/kg and xylazine 10 mg/kg (Rompun, Bayer AG, Leverkusen, Germany) were given intraperitoneally. As the anesthetics, ketamine (Ketalar, Pfizer, Turkey) 75 mg/kg and Xylazine 10 mg/kg (Rompun, Bayer AG, Leverkusen, Germany) were given intraperitoneally. For asepsis, polyvinylpyrolidone (polyvidon-iodine) 10% solution (PBS, Sigma Aldrich, St. Louis, MO, USA) containing penicillin-streptomycin, the tissue was cut into small pieces. After 1 hour of enzymatic degradation at 37°C with 5 ml Type I collagenase (1mg/mL, Sigma), 20–30 mL Low Glucose Dulbecco’s Modified Eagle’s Medium (L-DMEM, Gibco) with 1% penicillin-streptomycin (PS, Sigma), 1% L-glutamine (Sigma), and 10% fetal bovine serum (FBS, Gibco) were included. The mixture was centrifuged at 1.500 rpm for 10 minutes, and after removing the supernatant portion and by adding 15–20 mL of medium on the cell pellet, the centrifuge was continued for 10 min. at 1.200 rpm. After removing the supernatant portion, 12 mL of medium was added on the ADSC pellet and then, within a 75 cm² flask, incubated in a drying oven at 37°C with 5% CO₂. The ADSC cultures were used after the third passage.

Evaluation of the Adhesions and Tissue Sampling
On the 14th day, surgery was performed in rats under intraperitoneal anesthesia with ketamine (Ketalar, Pfizer, Turkey) 75 mg/kg and xylazine 10 mg/kg (Rompun, Bayer AG, Leverkusen, Germany). For the rats under anesthesia, to prevent the adhesion-caused injuries and to evaluate the adhesions on incision, laparotomy was performed via left paramedian incision. After the rating, tissue samples were taken from the cecum and adjacent region adhesions to perform the histopathologic and other examinations. The samples taken were divided into two equal parts. One part of the tissue samples was fixed with 1 cc physiological saline solution, and kept at −80°C for hydroxyproline determination. The remaining part was fixed in formaldehyde for a histopathological examination. All rats were sacrificed by investigators via cervical dislocation.

Histopathologic Analysis
Histopathologic analyses were performed by the same pathologist. The pieces were prepared in a paraffin block, and their thin sections were analyzed under the light microscope with hematoxylin–eosin staining; the images were recorded via the computer. The histopathologic staging was performed in accordance with the Ehrlich–Hunt model (16) (Table 1).

In this model, the assessment criteria were determined to be inflammatory cells, fibroblast, neovascularization, and the collagen level. The cellular and histopathologic scoring were semi-quantitatively determined to be at the fourth level. Separate calculations were performed for inflammatory cells, fibroblast proliferation, neovascularization, and the collagen deposition.

Immunohistochemical Analysis
For the immunohistochemical analysis, tissue samples were taken from all the rats in study groups and the control group. For the sections, the formalin fixation, paraffin treatment, and blocking were performed, and then, the immunohistochemical staining was carried out. In the tissues of different groups, the level of E-cadherin was semi-quantitatively determined (absent (0), slight (1): up to 20% positive; moderate (2): 21%–50% positive; potent (3): 51%–100%).

Tissue Hydroxyproline Level Determination
The hydroxyproline content of the tissues was determined using the Hydroxyproline Assay Kit from Sigma (Cat. No: MAK008). The assay method is based on the reaction of oxidized hydroxyproline with 4-dimethylaminobenzaldehyde (DMAB) to give a colorimetric product at 560 nm (1). To explain briefly, 100 µL of water was added on 10 mg of wet tissue, and homogenization was performed by a sonicator equipped with a microtip (Bandelin, Germany) at the amplitude of 25% (0.7 s on, 0.2 s off cycle). Sonication was continued until the tissue suspension became completely homogeneous, which usually took approximately 1–2 min of total sonication time. 100 µL of homogenized tissue sample was mixed with 100 µL of concentrated HCl (12 M), and
the mixture was hydrolyzed at 120 °C for 3 hours. Once the hydrolysis was complete, the samples were centrifuged at 13,000 g for 10 min, and 20 μL of each hydrolysate was transferred into the wells of a 96-well plate. The plate was incubated at 60 °C for 3–4 hours to dry the samples completely. This was followed by the addition of 100 μL chloramine T solution to each well and incubation for 5 min at the room temperature for the oxidation of hydroxyproline. Afterwards, 100 μL of the DMAB reagent was added into the wells, and the plate was incubated at 60°C for 90 min. A microplate reader (Multiskan GO, Thermofisher Scientific) was used to measure the absorbance at 560 nm immediately after incubation. The hydroxyproline standards with known concentration were also measured in a similar way. The amount of the hydroxyproline content was calculated using a standard curve that was generated from the absorbance values of standards. The results were expressed as micrograms of hydroxyproline per milligrams of wet tissue (μg/mg).

Statistical Analysis

For statistical analyses, Statistical Package for the Social Sciences (SPSS) version 13.0 (SPSS Inc.; Chicago, IL, USA). The

<table>
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<th>Table 1. Histologic Grading Scale*</th>
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<td>*Ehrlich–Hunt Model</td>
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<th>Table 2. Comparison of the Group Adhesion Scores</th>
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<td>General adhesion score</td>
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<td>6±1.1a</td>
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<tr>
<td>Adhesion severity</td>
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<td>Degree of adhesion</td>
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<td>Participation percentage</td>
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<td>*Kruskal–Wallis test: The results were given as the mean±standard deviation</td>
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<td>*Subgroup comparison: Statistically significant groups</td>
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<th>Table 3. Comparison of Histopathologic Findings and the Hydroxyproline Levels of Groups</th>
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<td>Group 1</td>
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<td>Inflammation</td>
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<td>Hydroxyproline (μg/mg)</td>
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<td>*Subgroup comparison: Statistically significant groups. Minimum (median, maximum)</td>
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<td>**ANOVA</td>
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<td>Group 1: Control; Group 2: Saline; Group 3: Adipose-derived mesenchymal stem cell</td>
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Figure 1. a-c. Macroscopic appearance of tissues. Control group: Intermediate increase adhesion is observed (a), Saline group: Intermediate increase adhesion is observed (b), Mesenchimal stem cell group: Increased adhesion is observed (c)
Kruskall–Wallis test was used in the semi-quantitative group comparisons. In the paired comparison, it was done using the Kruskal–Wallis (post hoc tests) and Sida–Dunn test. The analysis of variance was used in comparing the quantitative parameters. The Tukey test was employed for multiple comparisons. The results were given as the mean±standard deviation. The values p<0.05 were considered significant.

RESULTS

**Adhesion Scores**
The general adhesion score was 6±1.1 in Group 1, 4.4±1.1 in Group 2, and 7.5±1.2 in Group 3. The general adhesion score of Group 3 was found to be statistically significantly higher compared to the Group 1 and Group 2 (p=0.003). No statistically significant difference was found between the general adhesion scores of Group 1 and Group 2 (p>0.05; Table 2, Figure 1).

**Histopathologic Analysis Results**
In neovascularization and fibroblast intensity, no statistically significant difference was observed between the groups. In terms of collagen deposition, the statistically significant increase was found in Group 3 in proportion to Groups 1 and 2 (p=0.009). In inflammatory cell comparisons, a significant decrease was seen in Group 3 relative to Groups 1 and 2 (p=0.001). No statistically significant difference was found between Group 1 and Group 2 (p>0.05) (Table 3; Figure 2).

Figure 2. a-d. Histopathologic changes in Group 3. Grade 3 staining in Group 3: increase in the fibroblast intensity and a significant increase in neovascularization and the collagen fiber formation are observed (HE; X200) (a), Increase in collagen fiber and neovascularization in Group 3 (HE; X100) (b), Significant decrease in the number of inflammatory cells in Group 3 (HE; X100) (c), The pattern of Grade 3 staining with E-cadherin in Group 3 (immunohistochemistry; X400) (d)
Immunohistochemical Analysis Results
In comparisons performed between the groups, the pattern of positive staining with E-cadherin was found to be statistically higher in Group 3 compared with Groups 1 and 2 (p=0.003). No statistically significant difference was found between Group 1 and Group 2 (p>0.05) (Table 3).

Tissue Hydroxyproline Level Results
The hydroxyproline level was found to be 0.38±0.23 μg/mg in Group 1, 0.48±0.31 μg/mg in Group 2, and 0.40±0.06 μg/mg in Group 3. No statistically significant difference was found between the groups (p=0.69) (Table 3).

DISCUSSION
Intrapertitoneal adhesions are an important problem that surgeons have to cope with after the abdominal surgical procedures. The clinical situations caused by intraperitoneal adhesions and the complications, which develop due to intraperitoneal adhesions, lead to an increased mortality and morbidity (17). For this reason, preventing the intraperitoneal adhesions and dealing with the adhesions have constantly remained on the agenda of surgeons interested in the abdominal surgery. The studies have been carried out with many agents to prevent the adhesions. However, there is no consensus on the ideal agent to be used.

In the formation of postoperative intraperitoneal adhesions, damaged tissues and the fibrin deposits are responsible for the inflammation. The inflammatory process continues with the restoration of damaged tissues by the cells such as macrophages and fibroblast cells. The permanent fibrous adhesions are formed with collagen and other matrix proteins expressed from fibroblasts.

In the study by Sekiguchi et al. (18), it was reported that the bone marrow stem cells played an important role in the peritoneal restoration process and mesothelial remodeling. In that study, it was shown that the anti-inflammatory effect of the mesenchymal stem cells lasted for 14 days. In another study, the IL-10 cytokine having an inflammatory property has been found in the mesenchymal stem cell. The same study noted that the intraperitoneally injected mesenchymal stem cells suppressed the TGF-β1 expression and prevented the phosphorylation of Smad 2 caused by TGF-β1. These results indicate that the TGF-β1 signal was suppressed, and consequently, the mesenchymal stem cells showed an anti-inflammatory effect (19).

Although the tissue repair function and the immunoregulation of mesenchymal stem cells (MSCs) have been known for a long time, their exact mechanisms of action are still not clear. Recent findings indicate that an intravenous injection of MSCs may attenuate the peritoneal injury by repairing mesothelial cells, reducing inflammation and fibrosis. There remains a controversy on the application of MSCs in peritonitis, and peritoneal injury is still being explored. Mesenchymal stem cells segregate many growth factors. They are the strongest mediators of the HGF and BMP-7 anti-fibrotic cytokines. They play a preventive role in the formation of peritoneal fibrosis (20). The secretion of TSG-6 by MSCs makes a major contribution to the therapeutic benefits of MSCs. Moreover, when mesenchymal stem cells have been transplanted into injured tissues, an improvement was achieved in neovascularization and wound healing (21). ADSCs have been shown to have an anti-inflammatory effect by decreasing inflammation in the tissue (14, 20). While planning our study, we were focused on that ADSCs would have an anti-inflammatory effect, decrease the intra-abdominal adhesion, and might prevent it. Under the light of these clinical and experimental observations, ADSCs were used in the intra-abdominal adhesion model in our study.

In previous studies, it has been reported that mesenchymal stem cells increased the neovascularization and collagen level in tissues (18). Moreover, the increased collagen level may be correlated with an increase in the fibroblast intensity. Fibroblasts are responsible for the secretion of extracellular matrix proteins and collagen, and therefore, they play key role in wound healing. Parallel with the previous studies, we also found a significant increase in the ADSC group in proportion to the saline and control groups in terms of an increased collagen intensity. In our study, no statistically significant fibroblast intensity was found between the groups. However, fibroblast was relatively higher in the ADSC group. Mesenchymal stem cells can contribute to the regeneration in many organs (24). MSCs also increase the fibroblast regeneration. In our study, the increase in fibroblast intensity was thought to be correlated with the regenerative property of mesenchymal stem cells on the fibroblasts.

Hydroxyproline that constitutes the main component of collagen plays an important role in the collagen stabilization (25). The hydroxyproline level is used as an indicator of adhesion severity. An increased hydroxyproline level leads to an increase in peritoneal adhesion severity (26). In our study, the level of hydroxyproline was not found to be statistically significant in the groups. An increase in the general adhesion score in the ADSC group cannot be correlated with an increased hydroxyproline level.

Adipose derived mesenchymal stem cells produce too much cytokines, bioactive matters, extracellular matrix proteins, as well as adhesion molecules (20). In a study, it has been shown that the co-culture with ADSCs increased both the N-cadherin and E-cadherin expression (27). In our study, an increase was observed in the rate of positive staining with E-cadherin in the ADSCs group. E-cadherin is one of the molecules responsible for the intracellular adhesion and communication. An increase in the E-cadherin expression leads to an increase in the rate of cell-to-cell adhesion. Cellular adhesion molecules play an important role in the process of peritoneal adhesion formation.
The increase in E-cadherin expression may contribute to the increase in general adhesion score found in the ADSC group.

According to our hypothesis, it was thought that ADSCs would decrease the adhesion by exhibiting an anti-inflammatory effect and might prevent the adhesions. Despite an anti-inflammatory response, it was observed that ADSCs led to severe and strong intraperitoneal adhesions. The ADSCs group had the highest general adhesion score. Although ADSCs showed a desired anti-inflammatory efficiency, it was observed that they increased the adhesions. An increase in the E-cadherin expression and collagen can be held responsible for the adhesions.

Study Limitations
This study has several limitations. To the best of our knowledge, this is the first study in the literature that investigated the effects of ADSCs on the postoperative intraperitoneal adhesions formation. The ADSCs-associated histopathologic changes at the ultrastructural level need to be further analyzed. Therefore, further studies are needed to clarify the pathophysiological events. ADSCs were used 1 – 5x10^5 doses for different purposes and in different studies (28, 29). Although different time points could be considered, a single time point is used to evaluate the adhesion formation in this study. The vascular markers such as CD31 can be used for neovascularization. There are insufficient data on the rejection in allogeneic ADSCs. However, Kim et al. (30) studied immune reactions of allogenic adipogenic differentiated ADSCs. They showed that allogeneic adipogenic-differentiated ADSCs do not cause immunoreaction and maintain nonimmunogenicity. For this reason, they reported that allogeneic ADSCs could be used safely in recipients. Since all these findings were obtained in vitro, there is an urgent need in vivo studies for the immune reaction and rejection of allogeneic ADSCs. It is not known whether the allogenic ADSC used in our study caused rejection. Immunity can also be a response to the significant adhesion formation.

Adipose derived mesenchymal stem cells are not seen as an ideal agent for preventing the postoperative intraperitoneal adhesions. But, under the clinical conditions, where the formation of adhesions is desired, it can be used in the treatment such as closing the dead gaps via the formation of adhesion.

CONCLUSION
A severe adhesion was observed in the ADSCs-administered group. Also, the collagen intensity and E-cadherin expression increased in the ADSCs-administered group. The anti-inflammatory effect was also seen in the ADSCs group. The effect on the failure of adhesion improvement caused by an ADSCs intraperitoneal injection needs to be further analyzed. Further studies are needed to clarify the pathophysiological events.

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